

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/316452143>

# Rhizospheric Inoculation Influence on Seedling Growth, Development and Biomass Yield in *Oroxylum indicum* (L.) Benth. ex Kurz

Article · September 2015

CITATION

1

READS

51

2 authors:



**Chandrima Debi**

Forest Research Institute Dehradun

8 PUBLICATIONS 4 CITATIONS

[SEE PROFILE](#)



**Vipin Parkash**

Rain Forest Research Institute

57 PUBLICATIONS 335 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Root fungal endophytes and endomycorrhizal association in *Angelica glauca* and their role in accumulation of some bio-active compound/s [View project](#)



It is first of a kind comparison of the AMF associated with *Oroxylum indicum* of geographically two different regions from NE India. [View project](#)

# Rhizospheric Inoculation Influence on Seedling Growth, Development and Biomass Yield in *Oroxylum indicum* (L.) Benth. ex Kurz

Chandrima Debi<sup>1</sup>, Vipin Parkash<sup>2</sup>

<sup>1</sup>Ph.D. Scholar, Rain Forest Research Institute (ICFRE), Soil Microbiology Laboratory, SFM Division, Jorhat, Assam, India, 785001

<sup>2</sup>Scientist-E, Rain Forest Research Institute (ICFRE), Mycology and Soil Microbiology Research Laboratory, SFM Division, Jorhat, Assam, India, 785001

**Abstract:** *Oroxylum indicum* is a medicinally important forest tree species. Due to indiscriminate exploitation for medicinal purpose, habitat destruction, the natural population of *O. indicum* is reported endangered. Therefore an experiment was conducted to study the effect of bioinoculation on the growth and quality of the seedlings and conserve the plant species. The seedlings were inoculated with plant growth promoting microbes mainly, bacteria (B), fungus (F) and mycorrhiza (M), both alone and consortium. The total biomass was maximum ( $156.01 \pm 1.94$ ) in  $T_F$  treatment and minimum ( $118.78 \pm 0.06$ ) in Control treatment. The biovolume index was maximum ( $64.15 \pm 3.088$ ) in  $T_{M+B+F}$  and minimum ( $14.43 \pm 0.33$ ) in Control treatment. The Quality index (Qi) of the seedlings was maximum ( $1.041 \pm 0.089$ ) in  $T_B$  (seedlings treated with bacteria) while, Qi was minimum ( $0.761 \pm 0.090$ ) in Control treatment.

Key words: *Oroxylum indicum*, endangered, bioinoculation, biovolume index, quality index.

## 1. Introduction

*Oroxylum indicum* (L.) Benth. ex Kurz is a medicinally significant forest tree species, which is reported to be endangered and vulnerable in different parts of North east India as well as the entire Indian subcontinent. Problems associated with its natural propagation and indiscriminate exploitation for medicinal purpose has pushed *O. indicum* to the list of endangered plant species of India [1]. This plant is used in many ayurvedic preparations and widely used by people for health care. The important medicinal principles obtained from it are *Dashmula*, *Shyonaka patpak* and *Bruhat pancha mulayadi kwath* [2], *Chyawanprasha* [3] and is one of the best known health care products of Ayurveda. This tree has been harvested so heavily for medicinal purposes that its survival is in jeopardy.

The production, consumption and international trade of medicinal plants and phytomedicine (herbal medicine) is increasing. In order to satisfy growing market demands, there is need to develop new strategies for better yield and quality of medicinal plants, which can otherwise be achieved through various biotechnological methods. It may help in conserving many valuable tree species in the process and pave new vistas in forest biotechnology [4].

Plant growth is influenced by the presence of bacteria and fungi, and their interactions are particularly common in the rhizospheres of plants with high relative densities of microbes [5]. Microbial communities in the soil or rhizosphere contribute to plant growth by recycling nutrients and making them available [6], increasing root health through competition with root pathogens [7] or enhancing nutrient uptake [8].

Arbuscular mycorrhizal fungi (AMF) form symbiotic association which is ubiquitous and known to improve the nutritional status of the host plants by facilitating absorption of relatively immobile micronutrients such as Zn and Cu besides P through external mycelium that assists in nutrient transport [9,10,11,12,13,14]. AMF inoculation not only promotes the growth of medicinal plants but also improves the productivity and quantity of chemicals [15]. *Trichoderma* sp. also helps to mobilize and take up soil nutrients, which makes it more efficient and competitive than many other soil microbes [16]. Similarly, certain bacteria provide plants with growth promoting substances and play major role in phosphate solubilizing [17]. Phosphate solubilizing microorganisms are another sort of bio-fertilizers which have the ability to solubilize organic and inorganic phosphorus compounds by producing organic acid or phosphatase enzyme [18]. Mixed inoculation with diazotrophic bacteria and arbuscular-mycorrhizal fungi creates synergistic interactions that may result in a significant increase in growth, in the phosphorus content in plants, enhanced mycorrhizal infection and an enhancement in the uptake of mineral nutrients such as phosphorus, nitrogen, zinc, copper, and iron [19,20,21,22,23,24,25,26,27, 28].

The literature review revealed that bioinoculation studies has been mainly carried on various fields including the influence of bioinoculants on growth and mycorrhizal occurrence in the rhizosphere, plant growth stage, fertiliser management and bio-inoculation of arbuscular mycorrhizal fungi, impact of endomycorrhizal fungi and other bioinoculants on growth enhancement. There are many reports, projects and research work carried out on *Oroxylum indicum* such as propagation of *O. indicum*, through organogenesis [29] genetic diversity in *O. indicum*, by random amplified polymorphic DNA marker

Volume 5 Issue 9, September 2016

[www.ijsr.net](http://www.ijsr.net)

Licensed Under Creative Commons Attribution CC BY

[30], need and importance of conservation of endangered tree *O. indicum*, [31]. *In vitro* propagation of *O. indicum*: a medicinally important forest tree [32], antibacterial activity of stem bark extracts of *O. Indicum* [33], phytochemical and antimicrobial study of *O. indicum* [34], study on the secondary metabolites of *O. indicum* [35]. But there is no report on the effect of bioinoculation on growth and quality of the seedlings of *Oroxylum indicum* (L.) Benth. ex Kurz: Hence, the present study was undertaken.

## 2. Material and Methods

In order to analyse the the effect of bioinoculation on growth and quality of the seedlings of *O. indicum*, an experiment was setup in the nursery of Rain Forest Research Institute, Jorhat. The experiment was designed in Randomized Block Design (RBD), where three replications of each treatment were taken. Different treatments like single and combined/ synergistic/influential were applied for the present investigation. In control sets, no bioinoculant (inoculum) was added. Seedlings were raised in polyethene bags (30cm×50cm) containing 5 kg of substrates (sand and soil) in a ratio of 1:3. The inoculum was applied very close to the rhizosphere of the stumps at the depth of 5-10 cm. Three-stage inoculation was done e.g. at 0 day (1<sup>st</sup> stage, when the experiments will be initiated), 60 day (2<sup>nd</sup> stage) and 120 days (3<sup>rd</sup> stage). The inocula was added as per the previous studies conducted at concerned laboratory e.g. 10% w/w or w/v (1<sup>st</sup> stage), 20% w/w or w/v (2<sup>nd</sup> stage) 30% w/w or w/v (3<sup>rd</sup> stage) according to growing phases of seedlings. Data on plant growth like height, girth, the increment in diameter, was observed after definite intervals e.g. 90, 180, 270 days after inoculation (DAI). The effect of bio-inoculation on height, diameter and girth was recorded periodically and tabulated. The biomass, biovolume index and quality index of the seedlings was determined after 270 days of inoculation.

## 3. Biomass Estimation

Shoot biomass and Root biomass of a plant were calculated using the following formulae,

$$\text{Shoot biomass} = \frac{F_{w(s)} - D_{w(s)}}{F_{w(s)}} \times 100$$

$$\text{Root biomass} = \frac{F_{w(r)} - D_{w(r)}}{F_{w(r)}} \times 100$$

$$\text{Total biomass} = \text{Shoot biomass} + \text{Root biomass}$$

Where,  $F_{w(s)}$  = Fresh weight of shoot,  $F_{w(r)}$  = Fresh weight of root,  $D_{w(s)}$  = Dry weight of shoot,  $D_{w(r)}$  = Dry weight of root.

**Biovolume index:** - The biovolume index of the seedlings was calculated using the following formula [36, 37].

$$B_i = H \times D$$

Where  $B_i$  = Biovolume index,  $H$  = Height of seedlings in cm,  $D$  = Diameter of stem in mm/cm

**Quality index:** - Quality index to access the quality of seedlings was calculated using the following formula [36, 37].

$$Q_i = \frac{M_1}{\frac{H}{D} + M_2}$$

Where  $Q_i$  = Quality index,  $M_1$  = Seedlings whole biomass (shoot biomass + root biomass),  $M_2$  = Seedlings top biomass (except root biomass,  $H$  = Height of seedlings in,  $D$  = Diameter of stem in mm/cm

The results were analyzed statistically using standard error of mean and the test of significance [38].

## 4. Results

The effect of bioinoculation on the seedlings of *O. indicum* was studied and the data was tabulated accordingly. Table 1.1 represents the initial data of the height, diameter and girth of the seedlings of *Oroxylum indicum* before inoculation. T denotes treatment, M denotes Mycorrhiza, B = Bacteria (*Pseudomonas* sp.), F = Fungi (*Trichoderma harzianum*), ± SEM (Standard Error of mean), \* Average of Three replications

The initial height of the seedlings varied from 8.16±4.71 to 16.1±9.29 cms, the initial diameter varied between 0.233±0.017 to 0.439±0.041 cms and the initial girth varied from 0.733±0.053 to 1.380±0.226 cms. The data on the increase in height (cm), diameter and girth was recorded after 90 Days, 180 Days, and 270 days after inoculation. The data on the effect of bio-agents inoculation on height, diameter and girth of *Oroxylum indicum* after first stage inoculation 90 (DAI) of *Oroxylum indicum* was tabulated (Table 1.2). Table 1.3 shows the increase in height, diameter and girth of the seedlings due to bioinoculation. The maximum increase in height (11.533±3.066) in  $T_{M+B+F}$ , whereas, minimum (2.366±1.484) increase in height was recorded in treatment  $T_{M+B}$ . The increase in diameter after 90 days of inoculation was recorded maximum (0.12±0.0221) in  $T_{M+B+F}$  whereas minimum (0.040±0.016) was found in Control. The increase in girth was maximum (0.376±0.069) in  $T_{M+B+F}$  treatment, whereas, minimum (0.126±0.051) increase in girth was recorded in Control. Table 1.4 shows the effect of bioinoculation on height, diameter and girth of *Oroxylum indicum* after second stage inoculation i.e. 180 (DAI).  $T_{M+B+F}$  showed maximum height (46±1.00), whereas,  $T_{M+B}$  showed minimum height (24.06±0.52) after 180 days of inoculation. Diameter of the seedlings after 180 DAI shows maximum (0.59±0.064 and 0.59±0.048) diameter in  $T_B$  and  $T_F$  respectively. Control showed minimum (0.322±0.0319) diameter, while, girth was maximum (1.867±0.151) in  $T_F$  while it was minimum (1.011±0.100) in Control. The study of the effect of inoculation on increase in height, diameter and girth of *O. indicum* after second stage inoculation 180 (DAI) as tabulated in Table 1.5 reveals that the maximum increase in height was recorded in  $T_{M+B+F}$  while it was minimum in  $T_{M+B}$ . The maximum (0.242±0.058) increase in diameter of *O. indicum* was found in  $T_{F+B}$  while minimum increase of

diameter (0.088±0.015) was found in Control. The maximum (0.760±0.183) increase in girth was found in T<sub>F+B</sub> while it was minimum (0.277±0.048) in Control. After 270 days of inoculation, the effect on height, diameter and girth of *Oroxylum indicum* was recorded in Table 1.6. The height was maximum (77.83±4.15) in T<sub>M+B+F</sub> treatment, while it was minimum (39.2±3.90) in Control. Maximum (0.825±0.026) diameter was found in T<sub>M+B+F</sub> while minimum (0.377±0.029) diameter was recorded in Control. Table 1.7 shows the effect of bioinoculation on increase in height, diameter and girth on the seedlings of *O. indicum*. T<sub>M+B+F</sub> treatment showed maximum increase in height (65±3.09), whereas, minimum increase in height (28.93±3.61) was found in Control. T<sub>M+B+F</sub> showed maximum increase in diameter (0.404±0.058), whereas, T<sub>M+B</sub> treatment showed minimum increase in diameter (0.206±0.092). Maximum (1.598±0.068) increase in girth was recorded in T<sub>M+B+F</sub> treatment, whereas, minimum increase in girth (0.438±0.048) was recorded in Control.

Table 1.8 and Table 1.9 show the fresh and dry weight of the root and shoot of the seedlings of *O. indicum* respectively. It was found that the fresh weight of shoot (gm) was maximum (64.1±4.59) in case of T<sub>M+B+F</sub> while it was minimum in case of Control, the fresh weight of root (gm) was maximum (152.4±7.52) in T<sub>M+B+F</sub> while it was minimum (57.3±4.16) in Control. The total fresh weight (gm) was maximum (210.3±11.41) in case of T<sub>F</sub> while it was minimum (91.93±6.35) in case of Control. Similarly, the total dry weight (gm) of both root and shoot of the seedlings was taken. It was found that the dry weight of shoot was maximum (18.2±0.51) in T<sub>BF</sub>, while it was minimum (16.2±3.00) in case of T<sub>MF</sub>, whereas, the dry weight of root was maximum (26.66±1.16) in T<sub>M+B+F</sub>, while it was minimum (18.1±2.45) in case of T<sub>MB</sub>. The total dry weight (gm) was maximum (43.3±2.3) in case of T<sub>F</sub> while it was minimum (34.9±4.44) in case of T<sub>MB</sub>.

The biomass of the seedlings after 270 days of inoculation was calculated and tabulated in Table 1.10. The shoot biomass was maximum (73.92±1.77) in T<sub>M+B+F</sub> treatment and it was minimum (52.45±0.91) in Control. The root biomass was maximum (85.28±0.52) in T<sub>F</sub> treatment, whereas it was minimum (66.33±0.89) in Control. The total biomass was maximum (156.01±1.94) in T<sub>F</sub> treatment and minimum (118.78±0.06) in Control.

Table 1.11 shows the effect of bioinoculation on biovolume index. The initial biovolume index was maximum (6.75±0.003) in T<sub>M</sub> treatment and minimum (4.266±0.469) in Control (2.407±0.0003) before the initiation of experiment. The biovolume index was maximum (10.66±1.035) in T<sub>M+B+F</sub> and minimum (4.266±0.469) in Control after 90 days of inoculation. After 180 days of inoculation the biovolume index was again maximum (24.83±1.294) in T<sub>M+B+F</sub> and minimum (8.147±0.369) in Control. The biovolume index was maximum (64.15±3.088) in T<sub>M+B+F</sub> and minimum (14.43±0.33) in Control after 270 days of inoculation. Table 1.12 shows the Quality index (Qi) of seedlings of *O. indicum* after 270 DAI. The Qi was maximum (1.041±0.089) in T<sub>B</sub> while, Qi was minimum (0.761±0.090) in Control.

**Table 1.1:** Initial height, diameter and girth of the seedlings of *Oroxylum indicum* before inoculation 90(DAI)\*.

Treatments	Initial Height (cm)	Initial Diameter (cm)	Initial girth (cm)
Control	10±5.77	0.233±0.017	0.733±0.053
T <sub>M</sub>	16.1±9.29	0.418±0.02	1.313±0.151
T <sub>B</sub>	12.6±7.27	0.439±0.041	1.380±0.226
T <sub>F</sub>	11.5±6.63	0.386±0.017	1.214±0.093
T <sub>M+B</sub>	10.83±6.25	0.31±0.011	0.973±0.061
T <sub>M+F</sub>	9±5.19	0.351±0.048	1.102±0.262
T <sub>F+B</sub>	8.16±4.71	0.323±0.005	1.016±0.027
T <sub>M+B+F</sub>	15.5±7.89	0.316±0.004	0.993±0.025

Anova: F obtained value 89.22, p value 3.73, F<sub>critical</sub> value 3.008

**Table 1.2:** Effect of bioinoculation on seedling height, diameter and girth of *Oroxylum indicum* after first stage inoculation 90 (DAI)\*

Treatments	Height after 90 DAI	Diameter after 90 DAI	Girth after 90 DAI
Control	15.53±0.176	0.274±0.027	0.860±0.085
T <sub>M</sub>	18.46±2.37	0.495±0.031	1.554±0.098
T <sub>B</sub>	17.33±2.88	0.488±0.02	1.532±0.064
T <sub>F</sub>	20.03±2.161	0.429±0.01	1.349±0.054
T <sub>M+B</sub>	14.7±1.49	0.394±0.026	1.239±0.083
T <sub>M+F</sub>	13.73±2.96	0.423±0.056	1.328±0.178
T <sub>F+B</sub>	15.33±2.198	0.427±0.047	1.342±0.150
T <sub>M+B+F</sub>	24.36±1.70	0.436±0.218	1.370±0.068

**Table 1.3:** Effect of bioinoculation on increase in seedling height, diameter and girth of *Oroxylum indicum* after first stage inoculation 90 (DAI)\*

Treatment	Increase in height after 90 DAI	Increase in Diameter after 90 DAI	Increase in Girth after 90 DAI
Control	5.2±0.503	0.040±0.016	0.126±0.051
T <sub>M</sub>	2.433±0.578	0.076±0.019	0.24±0.060
T <sub>B</sub>	9±1.951	0.0483±0.023	0.151±0.074
T <sub>F</sub>	6.2±1.625	0.043±0.003	0.135±0.011
T <sub>M+B</sub>	2.366±1.484	0.084±0.036	0.265±0.116
T <sub>M+F</sub>	4.566±3.117	0.072±0.0105	0.226±0.033
T <sub>F+B</sub>	5.333±1.716	0.103±0.023	0.325±0.153
T <sub>M+B+F</sub>	11.533±3.066	0.12±0.0221	0.376±0.069

**Table 1.4:** Effect of bioinoculation on height, diameter and girth of *Oroxylum indicum* after second stage inoculation 180(DAI)\*

Treatment	Height after 180 DAI	Diameter after 180 DAI	Girth after 180DAI
Control	25.56±1.33	0.322±0.0319	1.011±0.100
T <sub>M</sub>	39.6±1.59	0.586±0.055	1.840±0.173
T <sub>B</sub>	35.16±6.99	0.59±0.064	1.856±0.202
T <sub>F</sub>	39.53±4.39	0.59±0.048	1.867±0.151
T <sub>M+B</sub>	24.06±0.52	0.420±0.060	1.320±0.191
T <sub>M+F</sub>	26.4±8.31	0.426±0.050	1.339±0.159
T <sub>F+B</sub>	29.6±2.94	0.565±0.053	1.776±0.168
T <sub>M+B+F</sub>	46±1.00	0.539±0.018	1.694±0.059

**Table 1.5:** Effect of bioinoculation on increase in height, diameter and girth of *Oroxylum indicum* after secondstage inoculation 180 (DAI)\*

Treatment	Increase in Height after 180 DAI	Increase in Diameter after 180 DAI	Increase in Girth after 180 DAI
Control	15.23±1.12	0.088±0.015	0.277±0.048
T <sub>M</sub>	23.56±2.90	0.167±0.273	0.526±0.086
T <sub>B</sub>	26.837.85	0.151±0.075	0.475±0.235
T <sub>F</sub>	25.7±4.80	0.208±0.031	0.653±0.098
T <sub>M+B</sub>	11.73±1.85	0.110±0.068	0.346±0.215
T <sub>M+F</sub>	17.23±5.39	0.075±0.002	0.236±0.008
T <sub>F+B</sub>	19.6±2.05	0.242±0.058	0.760±0.183
T <sub>M+B+F</sub>	33.16±2.36	0.223±0.015	0.700±0.047

**Table 1.6:** Effect of bioinoculation on height, diameter and girth of *Oroxylum indicum* after third stage inoculation 270 (DAI)\*

Treatment	Height after 270 DAI	Diameter after 270 DAI	Girth after 270DAI
Control	39.2±3.90	0.377±0.029	1.172±0.091
T <sub>M</sub>	57.66±8.17	0.701±0.095	2.203±0.299
T <sub>B</sub>	54.8±4.88	0.703±0.077	2.208±0.242
T <sub>F</sub>	61.1±7.129	0.744±0.040	2.336±0.127
T <sub>M+B</sub>	65.96±5.57	0.516±0.082	1.622±0.259
T <sub>M+F</sub>	52.6±17.26	0.575±0.057	1.807±0.179
T <sub>F+B</sub>	65.43±4.59	0.717±0.057	2.254±0.179
T <sub>M+B+F</sub>	77.83±4.15	0.825±0.026	2.592±0.824

**Table 1.7:** Effect of bioinoculation on increase in height, diameter and girth of *Oroxylum indicum* after third stage inoculation 270 (DAI)\*

Treatment	Increase in height after 270 DAI	Increase in Diameter after 270 DAI	Increase in Girth after 270 DAI
Control	28.93±3.61	0.281±0.049	0.438±0.048
T <sub>M</sub>	41.63±7.17	0.283±0.069	0.889±0.217
T <sub>B</sub>	46.46±5.58	0.263±0.102	0.827±0.322
T <sub>F</sub>	47.26±7.20	0.357±0.024	1.122±0.078
T <sub>M+B</sub>	53.62±5.07	0.206±0.092	0.649±0.289
T <sub>M+F</sub>	56.26±2.96	0.224±0.009	0.704±0.028
T <sub>F+B</sub>	58.53±2.54	0.394±0.062	1.237±0.195
T <sub>M+B+F</sub>	65±3.09	0.404±0.058	1.598±0.068

**Table 1.8:** Fresh weight of root and shoot of seedlings of *O. indicum* after 270 DAI\*

Treatment	Fresh weight shoot (gm)	Fresh weight root (gm)	Total fresh weight (gm)
Control	34.63±2.24	57.3±4.16	91.93±6.35
T <sub>M</sub>	50.03±4.70	127.33±5.42	177.36±10.13
T <sub>B</sub>	58±4.85	123.43±9.07	181.43±12.30
T <sub>F</sub>	57.9±5.91	152.4±7.52	210.3±11.41
T <sub>MB</sub>	59.03±8.66	103.8±4.29	162.86±4.83
T <sub>MF</sub>	53.66±11.24	82.76±5.55	136.43±16.36
T <sub>BF</sub>	45.33±1.89	113.2±2.99	158.53±4.87
T <sub>MBF</sub>	64.1±4.59	143.36±8.19	207.46±8.37

**Table 1.9:** Dry weight of root and shoot of seedlings of *O. indicum* after 270 DAI\*

Treatment	Dry weight shoot (gm)	Dry weight Root (gm)	Total dry weight (gm)
Control	16.46±1.14	19.26±1.28	35.73±2.37
T <sub>M</sub>	18.46±0.55	22.83±1.07	41.3±1.33
T <sub>B</sub>	17.66±1.49	20.43±3.25	38.1±3.13
T <sub>F</sub>	16.83±1.44	22.43±1.49	39.2±1.20
T <sub>MB</sub>	16.86±2.21	18.1±2.45	34.9±4.44
T <sub>MF</sub>	16.2±3.00	19.4±1.42	35.6±4.33
T <sub>BF</sub>	18.2±0.51	18.6±1.56	36.8±1.85
T <sub>MBF</sub>	16.7±1.51	26.66±1.16	43.3±2.3

**Table 1.10:** Biomass of seedlings of *O. indicum* after 270 DAI\*

Treatment	Shoot biomass	Root biomass	Total biomass
T <sub>c</sub>	52.45±0.91	66.33±0.89	118.78±0.06
T <sub>M</sub>	62.49±3.30	82.07±0.14	144.57±3.28
T <sub>B</sub>	68.99±4.29	83.38±2.53	152.38±1.75
T <sub>F</sub>	70.72±1.89	85.28±0.52	156.01±1.94
T <sub>MB</sub>	71.3±0.57	82.33±2.93	153.63±2.86
T <sub>MF</sub>	69.55±1.10	76.58±0.28	146.13±1.33
T <sub>BF</sub>	59.80±0.62	83.59±1.10	143.40±0.69
T <sub>MBF</sub>	73.92±1.77	81.35±0.52	155.28±2.04

**Table 1.11:** Biovolume index (Bi) of seedlings of *O. indicum*

Treatment	Biovolume initial	Biovolume after 90 DAI*	Biovolume after 180 DAI*	Biovolume after 270 DAI*
T <sub>c</sub>	2.407±0.0003	4.266±0.469	8.147±0.369	14.43±0.33
T <sub>M</sub>	6.75±0.003	9.118±1.277	23.38±3.061	39.12±3.339
T <sub>B</sub>	3.700±0.001	8.518±1.62	19.9±1.555	38.69±6.336
T <sub>F</sub>	5.369±0.001	8.675±1.288	23.35±2.409	45.68±6.397
T <sub>MB</sub>	3.807±0.0008	5.874±0.952	10.12±1.474	33.24±2.733
T <sub>MF</sub>	3.466±0.003	5.838±1.606	12±5.074	38.16±6.288
T <sub>BF</sub>	3.226±0.490	6.731±1.522	16.97±3.102	49.27±4.818
T <sub>MBF</sub>	4.047±0.0008	10.66±1.035	24.83±1.294	64.15±3.088

**Table 1.12:** Quality index (Qi) of seedlings of *O. indicum* after 270 DAI\*

Treatment	Quality index (Qi)
T <sub>c</sub>	0.761±0.090
T <sub>M</sub>	1.001±0.151
T <sub>B</sub>	1.041±0.089
T <sub>F</sub>	1.026±0.061
T <sub>MB</sub>	0.775±0.135
T <sub>MF</sub>	0.795±0.013
T <sub>BF</sub>	0.920±0.036
T <sub>MBF</sub>	0.923±0.039

## 5. Discussion

The growth and sustainability of a plant species is dependent on the biotic and abiotic factors associated with the particular plant species. Microorganisms play vital role in the cycling of the nutrients in the rhizosphere of the plant. Many plant growth promoting rhizobacteria, mycoflora and mycorrhizal fungi also stimulate plant through direct or indirect interactions with the plant roots. In the present study also results are same with

the findings of Rani et al. [39]. Gill and Singh [40]; Parkash and Aggarwal [41], Parkash et al.[42]. They reported that the mutualistic association was accounted for better colonization and plant growth due to interchange of carbon, phosphate and nitrogen between host fungi and bacteria. Thus, it can be conferred that a synergistic interaction of the above mentioned microorganisms have direct impact on the growth and development of the plant species. In the present investigation also combined synergistic treatment showed positive growth effect on *Oroxylum indicum* seedlings.

Due to indiscriminate collection, over exploitation, uprooting of whole plants bearing roots, this plant has become endangered and vulnerable in different parts of the Indian subcontinent. The existence of *O. indicum* in natural population is highly threatened and has been categorized as vulnerable by the government of India [43]. Local healers and traders are collecting this species from the wild which is causing a severe threat to the existence of this plant species due to poor seed viability [44]. Referring to the availability and various uses of the target plant species, it was essential to take up the concerned study to conserve the target plant species by modern biotechnological eco-friendly methods to produce healthy and quality stock of superior germplasm.

Authors are thankful to Rain Forest Research Institute (Indian Council of Forestry Research & Education), Jorhat, Assam for providing the all possible laboratory facilities and experimental field for carrying out the research work.

## References

- [1] Tiwari, S., Singh, K., Shah, P. 2007. *In vitro* Propagation of *Oroxylum indicum*-An Endangered Medicinal Tree. *Biotechnology*, 6(2): 299-301.
- [2] Yasodha, R., Ghosh, M., Barthwal, S., Gurumurthi, K., 2004. Importance of biotechnological research in tree species of Dashamula. *Indian For.*, 130(1): 79-108.
- [3] Ghate, V.S. 1999. Bruhat panchmula in ethno-medic and Ayurved, *J. Med. Arom. Plant Sci.*, 21, 1099-1110.
- [4] Dubey, N.K, Kumar, R. and Tripathi, P. (2004) Global promotion of herbal medicine: India's opportunity. *Curr. Sci.* 86(1): 37-41. *IRJP*, 4(1)
- [5] Berg G., Smalla K., 2009. Plant species and soil type cooperatively shape the structure and function of microbial communities in the rhizosphere. *FEMS Microbiol Ecol*, Apr; 68(1):1-13
- [6] Lynch, J.M., 1990. *The Rhizosphere*. Chichester, UK.
- [7] Weller, D.M., Raaijmakers, J.M., Gardener, B.B.M., Thomashow, L.S., 2002. Microbial populations responsible for specific soil suppressiveness to plant pathogens. *Annual Review of Phytopathology* 40, 309-348.
- [8] Smith, S.E., Read, D.J., 1997. *Mycorrhizal Symbiosis*. Academic Press, London
- [9] Li X.L., Marschner H, Romheld V. 1991. Acquisition of phosphorus and copper by VA-mycorrhizal hyphae and root to shoot transport in white clover. *Plant Soil* 136: 49-57.
- [10] Jakobsen I, Abbott L.K., Robson A.D. 1992 External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifolium subterraneum* L. 1. Spread of hyphae and phosphorus inflow into roots. *New Phytol* 120: 371-380.
- [11] Liu A, Hamel C, Hamilton R.I., Ma B.L., Smith D.L. (2000) Acquisition of Cu, Zn, Mn and Fe by mycorrhizal maize (*Zea mays* L.) grown in soil at different P and micronutrient levels. *Mycorrhiza* 9: 331-336.
- [12] Ryan M.H., Angus J.F. (2003) Arbuscular mycorrhizae in wheat and field pea crops on a low P soil: increased Zn uptake but no increase in P uptake or yield. *Plant Soil* 250: 225-239
- [13] Subramanian K.S., Tenshia V., Jayalakshmi K and Ramachandran V. (2009) Biochemical changes and zinc fractions in arbuscular mycorrhizal fungus (*Glomus intraradices*) inoculated and uninoculated soils under differential zinc fertilization. *Appl Soil Ecol* 43(1) pp. 32-39
- [14] Subramanian K.S., Charest C. (1999) Acquisition of N by external hyphae of an arbuscular mycorrhizal fungus and its impact on physiological responses in maize under drought-stressed and well-watered conditions. *Mycorrhiza* 9, 69-75.
- [15] Karthikeyan B, Joe M.M., Jaleel C.A. 2009. Response of some medicinal plants to vesicular-arbuscular mycorrhizal inoculations. *Journal of Scientific Research* 1(2):381-386
- [16] Vinale F., Sivasithamparan K., Ghisalberti E.L., Marra R., Woo S.L., Lorito M. *Trichoderma*-plant-pathogen interactions. *Soil Biol Biochem* 2008; 40: 1-10.
- [17] Abou-Aly H.E., Mady M.A., Moussa S.A.M. 2006. Interaction effect between phosphate dissolving microorganisms and boron on growth, endogenous phytohormones and yield of squash (*Cucurbita pepo* L.). The First Scientific Conference of the Agriculture Chemistry and Environment Society, Cairo, Egypt.
- [18] Rashid M., Khalil S Ayub N, Alam S, Latif F. 2004. Organic Acid Production and Phosphate Solubilization by Phosphate Solubilizing Microorganism (PSM) under in vitro Conditions. *Pakistan Journal of Biology Science* 7, 187- 196.
- [19] Al-Nahidh, S. and Gomah, A.H.M. 1991. Response of wheat to dual inoculation with VA-mycorrhiza and *Azospirillum*, fertilized with NPK and irrigated with sewage effluent. *Arid Soil Res. Rehabil.*, 5: 83-96.
- [20] Barea, J.M. 1997. Mycorrhiza/bacteria interactions on plant growth promotion. In: *Plant Growth-Promoting Rhizobacteria-present status and future prospects*. Ogoshi, A., Kobayashi, K., Homma, Y., Kodama, F., Kondo, N. and Akino, S. (eds.), pp. 150-158, Faculty of Agriculture, Hokkaido University, Sapporo, Japan.
- [21] Garbaye, L. 1994. Helper bacteria: a new dimension to the mycorrhizal symbiosis. *New Phytol.*, 128:197-210.
- [22] Isopi, R., Fabbri, P., Del Gallo, M. and Puppi, G. 1995. Dual inoculation of sorghum bicolor (L.) Moench

- ssp. bicolor with vesicular arbuscular mycorrhizas and *Acetobacterdiazotrophicus*. Symbiosis, 18: 43-55.
- [23] Li, C.Y. and Huang, L.L. 1987. Nitrogen fixing (acetylene reducing) bacteria associated with ectomycorrhizas of Douglas -fir. Plant Soil, 98: 425-428.
- [24] Li, C.Y., Massicote, H.B. and Moore, L.V.H. 1992. Nitrogen-fixing *Bacillus* sp. associated with Douglas -fir tuberculateectomycorrhizae. Plant Soil, 140: 35-40.
- [25] Linderman, R.G. 1992. Vesiculararbuscularmycorrhizae and soil microbial interactions. In: Mycorrhizae in sustainable agriculture, Bethlenfalvay, G. J. and Linderman, R.G. (eds.), pp. 45-70, ASA Special Publication Number 54. American Society of Agronomy, Inc., Crop Science Society of America, Inc., Soil Science Society of America, Inc. Madison, USA.
- [26] Linderman, R.G. and Paulitz, T.C. 1990. Mycorrhizal-rhizobacterial interactions. In Biological control of soil-borne plant pathogens, Hornby, D. (ed.), pp. 261-283, CAB International, Wallington, UK.
- [27] Rózycki, H., Kampert, M., Strzelczyk, E., Li, C.Y. and Perry, D.A. 1994. Effect of different soil bacteria on mycorrhizae formation in scots pine (*Pinus sylvestris*L.)- *in vitro* studies. Folia Forestalia Polonica, 36: 91-102.
- [28] Singh, C.S., Amawate, J.S., Tyagi, S.P., Kapoor, A. 1990. Interaction effect of *Glomus fasciculatum* and *Azospirillum brasilense* on yields of various genotypes of wheat (*Triticumaestivum*) in pots. Z. Mikrobiol., 145: 203-208.
- [29] Jasrai, Y.T., Thaker, K.N., Parmar, V.R. 2013. Propagation of *Oroxylum indicum* (L.) Vent, a Vulnerable Medicinal Tree through Organogenesis. Plant Tissue Cult. & Biotech., 23(1): 127 - 132.
- [30] Jayaram, K., Prasad, M.N. 2008. Genetic diversity in *Oroxylum indicum* (L.) Vent. (Bignoniaceae), a vulnerable medicinal plant by random amplified polymorphic DNA marker. Afr. J. Biotech., 7:254-62.
- [31] Najar, Z.A. and Agnihotri, S. 2012. Need and Importance of Conservation of Endangered Tree *Oroxylum indicum* (Linn.) Vent.. Asian Journal of Plant Science and Research, 2(3): 220-223.
- [32] Dalal, N.V., Rai, V.R. 2004. *In vitro* propagation of *Oroxylum indicum* Vent. a medicinally important forest tree. J. For. Res., 9:61-65.
- [33] Samatha, T., Sampath, A., Sujatha, K. and Rama, S.N. 2013. Antibacterial Activity of Stem Bark Extracts of *Oroxylum indicum* an Endangered Ethnomedicinal Forest Tree, IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS), 7(1): 24-28.
- [34] Radhika, L.G., Meena, C.V., Peter, S., Rajesh, K.S., Rosamma, M.P. 2011. Phytochemical and antimicrobial study of *Oroxylum indicum*. Ancient Sci. Life, 30, 114-120.
- [35] Chen, L.J., Games, D.E., Jones, J. 2003. Isolation and identification of four flavonoids constituents from the seeds of *Oroxylum indicum* by high-speed counter-current chromatography. J. Chromatograph A., 988:95-105.
- [36] Parkash, V. Aggarwal, A. and Bipasha, 2011a. Rhizospheric effect of vesicular arbuscular mycorrhizal inoculation on biomass production of *Rutagraveolens*L.: A potential medicinal and aromatic herb. Journal of Plant Nutrition, 34( 9): 1386-96.
- [37] Hatchell, G. E. 1985. Biomass research. Proc. Third biennial Southern Silvicultural Res. Conf. Atlanta, Georgia, November, 7-8, 1984. General Technical Report, Southern Forest Experiments, Station, USDA Forest Service. 54: 395-402.
- [38] Pagano, R.R. 2000. Understanding Statistics in the Behavioural Sciences. Pub. Wadsworth Thompson Learning, Belmont, CA.
- [39] Rani, P., Aggarwal, A. and Mehrotra R.S. 1999. Growth responses in *Acacia nilotica* inoculated with VAM fungus *Glomus mosseae*, *Rhizobium* sp. and *Trichoderma harzianum*. Indian Phytopath., 52 (2) : 151-153.
- [40] Gill, T.S. and Singh R.S. 2002. Effect of *Glomus fasciculatum* and *Rhizobium* inoculation on VA mycorrhizal colonization and plant growth of chickpea. J. Mycol. Pl. Pathol., 32(2): 162-167.
- [41] Parkash, V. and Aggarwal, A. 2009. Diversity of endomycorrhizal fungi and its synergistic effect on growth of *Acacia catechu* Willd. Journal of Forest Science, 55(10): 461-468.
- [42] Parkash, V., Sharma, S. and Aggarwal, A. 2011b. Symbiotic and synergistic efficacy of endomycorrhizae with *Dendrocalamus strictus* L. Plant Soil Environment, 57(10): 447-451.
- [43] Ravikumar, K. and Ved, D.K. 2000. 100 Red listed medicinal plants of conservation concern in Southern India. Foundation for Revitalization of Local Health Traditions, Bangalore. India.
- [44] Chandrima Debi and Vipin Parkash (2015). Seed source and habitat variation affect seed germination in *Oroxylum indicum* (L.) Benth. ex Kurz: An important threatened medicinal tree. International Journal of Life Sciences and Technology, Volume 8(1):1-9.

### Author Profile



**Dr. Vipin Parkash** is serving as Scientist-E in the Indian Council of Forestry Research & Education at Rain Forest Research Institute, Jorhat, Assam. He has significantly contributed in this field and is expert in Mycology and Plant Pathology, Endomycorrhiza (AM fungi), Bio-control of soil borne pathogens, Soil Microbiology, Biological screening of compounds and secondary metabolites production and Plant Taxonomy & Systematics. He has more than 75 research publications in national as well as international journals having impact factors to his credit. He has also authored 4 books, guided 5 PhD students, 5 M.Phil. students and more than 20 post graduate students. He has also got 3 national and 1 international awards for outstanding scientific contribution and also discovered and described two new fungal species, i.e. *Lysurus habungianus* sp. nov., *Gelatinomyces conus* sp. nov.



**Chandrima Debi** is pursuing her Ph.D. research programme under the guidance and supervision of Dr. Vipin Parkash and doing research on influence of bioinoculation on the biotization and accumulation of bioactive compounds on medicinal plants.